NERC Open Research Archive



Article (refereed) - postprint

Woodcock, B.A.; Bullock, J.M.; Shore, R.F.; Heard, M.S.; Pereira, M.G.; Redhead, J.; Ridding, L.; Dean, H.; Sleep, D.; Henrys, P.; Peyton, J.; Hulmes, S.; Hulmes, L.; Sárospataki, M.; Saure, C.; Edwards, M.; Genersch, E.; Knäbe, S.; Pywell, R.F.. 2017. Country-specific effects of neonicotinoid pesticides on honey bees and wild bees. *Science*, 356 (6345). 1393-1395. 10.1126/science.aaa1190

2017 @ The Authors

This version available http://nora.nerc.ac.uk/518181/

NERC has developed NORA to enable users to access research outputs wholly or partially funded by NERC. Copyright and other rights for material on this site are retained by the rights owners. Users should read the terms and conditions of use of this material at http://nora.nerc.ac.uk/policies.html#access

This document is the author's final manuscript version of the journal article, incorporating any revisions agreed during the peer review process. Some differences between this and the publisher's version remain. You are advised to consult the publisher's version if you wish to cite from this article.

The definitive version is available at http://science.sciencemag.org/

Contact CEH NORA team at noraceh@ceh.ac.uk

The NERC and CEH trademarks and logos ('the Trademarks') are registered trademarks of NERC in the UK and other countries, and may not be used without the prior written consent of the Trademark owner.

1	Country-specific effects of neonicotinoid pesticides on noneybees and wild bees
2	
3	Authors: B.A.Woodcock ^{1*} , J.M.Bullock ¹ , R.F.Shore ² , M.S.Heard ¹ , M.G.Pereira ² , J.Redhead ¹ ,
4	L. Ridding ¹ , H.Dean ¹ , D.Sleep ² , P.Henrys ² , J.Peyton ¹ , S.Hulmes ¹ , L.Hulmes ¹ , M.Sárospataki ³ ,
5	C.Saure ⁴ , M.Edwards ⁵ , E.Genersch ⁶ , S.Knäbe ⁷ & R.F.Pywell ¹
6	
7	Affiliations:
8	¹ NERC Centre for Ecology & Hydrology, Oxfordshire OX10 8BB, UK.
9	² NERC CEH, Lancaster Environment Centre, Lancaster LA1 4AP, UK.
10	³ Szent-István University, 2103 Gödöllö, Hungary.
11	⁴ Am-Heidehof 44, 14163 Berlin, Germany.
12	⁵ Leaside, Carron Lane, West Sussex GU29_9LB, UK
13	⁶ Institute for Bee Research, 16540 Hohen-Neuendorf, Germany
14	⁷ Eurofins, Ecotox-GmbH, 75223 Niefern-Öoschelbronn, Germany
15	*Correspondence: B.A.Woodcock (<u>bawood@ceh.ac.uk</u>)
16	
17	Abstract:
18	Neonicotinoid seed dressings have caused concern world-wide. We use large field
19	experiments to assess effects of neonicotinoid-treated crops on three bee species across
20	three countries (Hungary, Germany and the UK). Winter-sown oilseed rape was grown
21	commercially with either seed coatings containing neonicotinoids (clothianidin or
22	thiamethoxam) or no seed treatment (control). For honeybee we found both negative
23	(Hungary and UK) and positive (Germany) effects during crop flowering. In Hungary,
24	negative effects on honeybees (associated with clothianidin) persisted over winter and
25	resulted in smaller colonies in the following spring (24% declines). In wild bees (Bombus
26	terrestris and Osmia bicornis), reproduction was negatively correlated with neonicotinoid
27	residues. These findings point to neonicotinoids causing a reduced capacity of bee species to
28	establish new nonulations in the year following exposure.

One Sentence Summary:

such effects differ between countries.

Honeybee and wild bee exposure to neonicotinoid pesticides reduces their ability to establish

32 populations.

Main Text:

Global declines in honeybees and wild bees have been linked to pathogens, climate change, habitat fragmentation and pesticide use (1-3). The potential threat from neonicotinoid seed coatings applied to flowering crops has been the subject of considerable debate (4-9). Neonicotinoids have been shown to increase mortality in honeybees by impairing their homing ability (4) and to reduce the reproductive success of bumblebees (5, 8, 10) and solitary bees (8, 11), while other studies have identified no effects (8, 12, 13). There is limited information from replicated studies on longer-term survival of honeybee colonies following exposure (see (12)). Landscape-scale experiments under real world agricultural conditions are needed to integrate spatial, temporal and species-specific variation to understand the impacts of neonicotinoids on bees (8, 12, 14-16). Such studies should explore the impacts of different neonicotinoid formulations, land use and regional climate. In a large-scale experiment spanning three European countries, we tested the hypotheses that: (i) exposure to seed treatments containing neonicotinoids affected the reproductive potential of managed and wild bee species and (ii) if

At each of 33 sites (Germany=9, Hungary=12, UK=12) an average of 63.1 ha (SE±2.8 ha) of winter-sown oilseed rape (OSR) was established in 2014 (Fig.1 & S1, Table S1). We clustered sites into triplets (>3.2 km between sites) and randomly allocated sites to one of three treatments: 1) Clothianidin applied at 11.86-18.05 g ha⁻¹ a.i. with a fungicide (Thriam and prochloraz) and non-systemic pyrethroid (beta-cyfluthrin) (trade name Modesto); 2) Thiamethoxam applied at 10.07-11.14 g ha⁻¹ a.i. and combined with the fungicides fludioxonil and metalaxyl-M (trade name Cruiser); 3) Control oilseed rape receiving a commercial fungicide (Thriam and Dimethomorph in Germany & Hungary, Thriam and Prochloraz in the UK), but no neonicotinoid seed treatment. All treatments received typical commercial inputs of pesticide (e.g. Lambda-cyhalothrin) and fertilizer, with these standardized across a triplet. Standardized colonies of honeybees (*Apis mellifera*) and wild bees (bumblebee *Bombus terrestris* and solitary bee *Osmia bicornis*) were introduced to each site. For honeybees we quantified the impacts of

- the treatments on colony viability during the crop flowering period and in the year following
- 62 exposure (hive survival and overwintering worker, brood and storage cell numbers).
- Overwintering fitness defines the multi-year persistence of honeybees. For *B. terrestris* we
- 64 measured impacts on within-year reproductive output (colony weight gain, and worker, queen
- and drone production) and for *O. bicornis* the number of reproductive cells produced (Table S2).
- Neonicotinoids can be persistent and widespread in agro-ecosystems (17, 18), so we quantified
- residues both in the nests of bee species and those expressed in the crop.

We found that neonicotinoid seed treatment affected the inter-annual viability of honeybee colonies following the winter period in a country-specific manner. In Hungary worker numbers were 24% lower where clothianidin was compared to the control (treatment×country: $\chi^2_6=1.47$, p=0.01, explained variance=59.4%; Fig.2), with no significant effect of thiamethoxam. Clothianidin was more likely to be expressed in the crop where it was applied as a seed treatment, which identified a mechanism of exposure to the bees ($\chi^2_2=6.46$, p=0.04), but this was not so for thiamethoxam (Table S3). In the UK high hive mortality precluded a formal statistical analyses of overwintering worker numbers. However, median worker numbers were zero for all four clothianidin-treated sites, but above zero for two of the control and one of the thiamethoxam

sites (Table S2; Fig.2). Worker numbers following the winter in Germany showed no treatment

effect (Table S4). Overwintering honeybee brood, stored hive products (pollen and nectar) and

the likelihood of hives surviving the winter were not affected by seed treatments (Table S3).

Neither *B. terrestris* queen nor *O. bicornis* egg cell production were directly affected by the seed treatments or its interaction with country (Table S5). However, they were negatively correlated with peak (χ^2_1 =2.09, p=0.03, explained variance=13.5%; Fig.3a) and median (χ^2_1 =4.34, p=0.04, explained variance=0.8%; Fig.3b) neonicotinoid nest residues (combined clothianidin, thiamethoxam and imidacloprid). Imidacloprid was not applied as part of the study and its presence is most likely a result of environmental contamination from previous widespread agronomic use (*17*, *18*). Residues of neonicotinoids detected in stored hive products did not differ in response to seed treatments for any bee species (Table S6). This may be due to the amalgamation of stored hive products at the site level for residue analysis, which may have obscured within site heterogeneity in residues. The negative correlation for *B. terrestris* queen production remained significant when we excluded sites with imidacloprid residues (χ^2_1 =2.14,

p=0.02), although this was not the case for O. bicornis (χ^2 ₁=0.05, p=0.81). Country-specific

responses to neonicotinoid seed treatment were found for *B. terrestris* drone production, with positive and negative effects from exposure to thiamethoxam in Germany and the UK respectively (treatment×country: χ^2_6 =13.1, p=0.04, explained variance=13.6%; Fig.2).

92

93

94

95

96

97

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115116

117

118

119

120

121

We also found seed treatment effects during the crop flowering period that lasted between 3 to 6 weeks (Table S4 & 5). Significant interactions between seed treatment and country were identified for peak worker (χ^2_6 =16.6, p<0.01, explained variance=45.3%), egg cell (χ^2_6 =4.13, p=0.01, explained variance=49.9%) and combined pollen and nectar storage cell (χ^2_6 =40.5, p<0.001, explained variance=53.6%) numbers. These responses describe within-year colony performance. Neonicotinoid exposure resulted in both negative (Hungary and UK) and positive (Germany) effects on colony size (see Fig.2; pairwise treatment comparison given in Table S4 & 5). *Bombus terrestris* worker and peak colony weight showed no seed treatment response.

Our quantification of neonicotinoid effects on the inter-annual viability of honeybees and wild bee populations represents a fundamental advance in our understanding of the impacts of these pesticides. For solitary bees and bumblebees (queen production) neonicotinoid impacts were associated with the residues found in nests rather than the experimental seed treatments. For B. terrestris the few treatment effects and the presence of imidacloprid in stored pollen and nectar (Table S7-S9) suggests that negative impacts of neonicotinoids may be driven by persistence of residues in the wider landscape, rather than current management alone (18, 19). The EU moratorium meant that no neonicotinoids were applied to oilseed in the surrounding landscapes during the experiment, so such residues may originate from previous agricultural use leading to expression in non-target plants (17-19), guttation fluids or contaminated water (19, 20). While the reproductive potential of O. bicornis was also negatively affected by neonicotinoid residues in nests, the explained variation of these effects was small. However, a failure to detect small population changes may be due to limited experimental replication restricting statistical power. Our results suggest that even if their use were to be restricted, as in the recent EU moratorium, continued exposure to neonicotinoid residues resulting from their previous widespread use has the potential to impact negatively wild bee persistence in agricultural landscapes (14, 18, 19).

Taken together, our results suggest that exposure to neonicotinoid seed treatments can have negative effects on the inter-annual reproductive potential of both wild and managed bees,

- but that these effects are not consistent across countries. The country-specific responses of
- honeybees and bumblebees strongly suggests that the effects of neonicotinoids are a product of
- interacting factors (20-23). This study has identified between country differences in the use of
- oilseed rape crop as a forage resources by bees (affecting exposure to crop residues) and
- incidence of disease within hives. Both factors were higher for Hungarian and UK honeybees
- 127 (Table S10 & S11). Overall neonicotinoid residues were detected infrequently and rarely
- exceeded 1.5 ng g⁻¹ w/w. As such, direct mortality effects caused by exposure to high
- concentrations of neonicotinoids are likely to be rare (Table S12). However, our results suggest
- that exposure to low levels of neonicotinoids may cause reductions in hive fitness that are
- influenced by a number of interacting environmental factors. Such interacting environmental
- factors can amplify the impact of honeybee worker losses (e.g. through sub-lethal toxicity
- effects) and reduce longer-term colony viability (4, 16). Importantly, our common experimental
- approach applied across three countries revealed varying impacts and may explain the
- inconsistent results of previous studies conducted in single countries or at few sites (4, 5, 8, 12,
- 136 *13*, *15*).

138

References and Notes:

- 1. A. J. Vanbergen, Insect Pollinators Initiative, Threats to an ecosystem service: pressures on pollinators. *Front. Ecol. Environ.* **11**, 251 (2013).
- 2. S. G. Potts *et al.*, Global pollinator declines: trends, impacts and drivers. *TREE* **25**, 345 (2010).
- R. Winfree, R. Aguilar, D. P. Vázquez, G. Lebuhn, M. A. Aizen, A meta-analysis of bees' responses to anthropogenic disturbance. *Ecology* **90**, 2068 (2009).
- M. Henry *et al.*, A common pesticide decreases foraging success and survival in honey bees.
 Science 336, 348 (2012).
- P. R. Whitehorn, S. O'Connor, F. L. Wäckers, D. Goulson, Neonicotinoid pesticide reduces Bumble Bee colony growth and queen production. *Science* **336**, 351 (2012).
- J. E. Cresswell *et al.*, Differential sensitivity of honey bees and bumble bees to a dietary insecticide (imidacloprid). *Zoology* **115**, 365 (2012).
- 150 7. B. A. Woodcock *et al.*, Impacts of neonicotinoid use on long-term population changes in wild bees in England *Nat. Comm.* 7, 12459 (2016).
- 152 8. M. Rundlöf *et al.*, Seed coating with a neonicotinoid insecticide negatively affects wild bees.

 153 *Nature* **521**, 77 (2015).

- 9. G. E. Budge *et al.*, Evidence for pollinator cost and farming benefits of neonicotinoid seed coatings on oilseed rape. *Sci. Report.* **5**, 12547 (2015).
- D. Goulson, Neonicotinoids impact bumblebee colony fitness in the field; a reanalysis of the UK's Food & Environment Research Agency 2012 experiment. *PeerJ* 3, e854 (2015).
- 158 11. C. Sandrock *et al.*, Sublethal neonicotinoid insecticide exposure reduces solitary bee reproductive success. *Agric. Forst. Entomol.* **16**, 119 (2014).
- 160 12. G. C. Cutler, C. D. Scott-Dupree, M. Sultan, A. D. McFarlane, L. Brewer, A large-scale field study examining effects of exposure to clothianidin seed-treated canola on honey bee colony health, development, and overwintering success. *Peerj* 2, 2167 (2014).
- 163 13. G. C. Cutler, C. D. Scott-Dupree, A field study examining the effects of exposure to neonicotinoid seed-treated corn on commercial bumble bee colonies. *Ecotox.* **23**, 1755 (2014).
- 165 14. B. A. Woodcock *et al.*, Replication, effect sizes and identifying the biological impacts of pesticides on bees under field conditions. *J. Appl. Ecol.* **53**, 1358 (2016).
- 15. E. Pilling, P. Campbell, M. Coulson, N. Ruddle, I. Tornier, A Four-Year Field Program

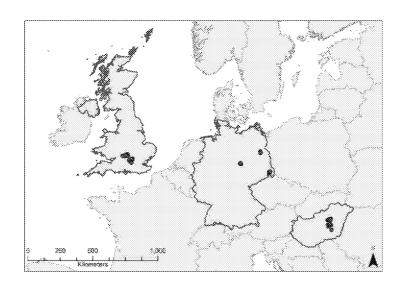
 Investigating Long-Term Effects of Repeated Exposure of Honey Bee Colonies to Flowering

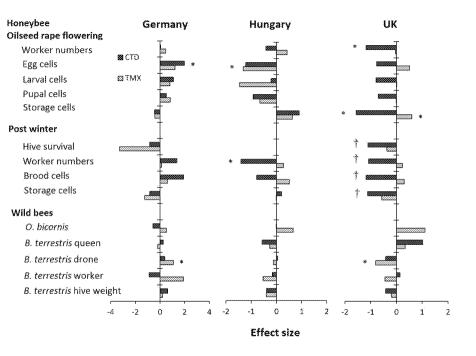
 Crops Treated with Thiamethoxam. *PLoS ONE* **8**, e77193 (2013).
- 170 16. M. Henry *et al.*, Reconciling laboratory and field assessments of neonicotinoid toxicity to honeybees. *Proc. R. Soc. Lond. Ser. B.* **282**, (2015).
- 172 A. Jones, P. Harrington, G. Turnbull, Neonicotinoid concentrations in arable soils after seed treatment applications in preceding years. *Pest Manag. Sci.* **70**, 1780 (2014).
- 174 18. C. Botías *et al.*, Neonicotinoid residues in wildflowers, a potential route of chronic exposure for bees. *Environ. Sci. Tech.* **49**, 12731 (2015).
- 176 19. D. Goulson, An overview of the environmental risks posed by neonicotinoid insecticides. *J. Appl. Ecol.* **50**, 977 (2013).
- 178 20. A. Fairbrother, J. Purdy, T. Anderson, R. Fell, Risks of neonicotinoid insecticides to honeybees.

 179 Environ. Toxicol. Chem. 33, 719 (2014).
- FERA, Neonicotinoid Pesticides and Bees. Report to Syngenta Ltd. *The Food and Environment Research Agency* (2013).
- F. Sanchez-Bayo *et al.*, Are bee diseases linked to pesticides? A brief review. *Environ. Int.* **89-90**, 7 (2016).
- 184 23. C. R. Archer, C. W. W. Pirk, G. A. Wright, S. W. Nicolson, Nutrition affects survival in African honeybees exposed to interacting stressors. *Funct. Ecol.* **28**, 913 (2014).
- 186 24. European Commission. (EC, http://ec.europa.eu/eurostat/web/agriculture/data/database) (2016).
- A. Imdorf, G. Buehlmann, L. Gerig, V. Kilchenmann, H. Wille, A Test of the method of estimation of brood areas and number of worker bees in free-flying colonies. *Apidologie* **18**, 137 (1987).

190 191	26.	D. Kleijn et al., Delivery of crop pollination services is an insufficient argument for wild pollinator conservation. <i>Nat. Commun.</i> 6 , 7414 (2015).	
192 193	27.	B. Magnusson, T. Näykki, H. Hovind, K. M., Nordtest report 537. Handbook for calculation of measurement uncertainty, (Nordic Innovation, Norway, 2012).	
194 195	28.	T. Blacquiere, G. Smagghe, C. A. M. van Gestel, V. Mommaerts, Neonicotinoids in bees: a review on concentrations, side-effects and risk assessment. <i>Ecotox.</i> 21 , 973 (2012).	
196 197	29.	EFSA, Conclusion on the peer review of the pesticide risk assessment for bees for the active substance clothianidin/thiamethoxam/imidacloprid, <i>EFSA</i> (2013).	
198 199	30.	R Core Development Team, R: Version 3.2.1 A Language and Environment for Statistical Computing. R Foundation for Statistical Computing. URL hhtp://cran.r-project.org. (2015).	
200			
201	Ackı	nowledgments: Data in supporting online material. Funded by Syngenta Ltd. and Bayer	
202		CropScience (P.Campbell, M.Miles, C.Maus, D.Holah, M.Coulson). Wild pollinator	
203		work supported by NERC CEH National Capability funding (NEC05829). Thanks to	
204		K.Jaekel, P.Fisher, M.Nowakowski, R.Hails, P.Scrimshaw, N.Mitschunas, P.Nuttall,	
205		M.McCracken, S.Ball, J.Webb, B.Sutherland, R.Freckleton, T.Tscharntke, J.Memmott,	
206		K.Norris, Brigitta Raffa and Dóra Vaskor.	
207	Cum	Jamontany content	
208		olementary content	
209	References (24-30)		
210	Materials and methods		
211	Figs.S1-2		
212	Table	es S1-12	
213			
214	Fig.1	. Location of the 33 experimental sites in the UK, Hungary and Germany. See Fig S2	
215	for a	diagrammatic representation of the experimental setup.	
216			
217	Fig.2	. Summary effect sizes for the response of honeybees and wild bees to the	
218	neon	icotinoid seed treatments. An effect size represents the difference between the mean	
219	population response for a given seed treatment and the control within a country, with this		
220	diffe	rence divided by the pooled standard deviation. Where: * indicates a significant differences	
221	betw	een the control and seed treatment (either TMX=thiamethoxam, CTD=clothianidin)	
222	deter	mined from the predicted marginal means of the model ' $v \sim seed treatment*country +$	

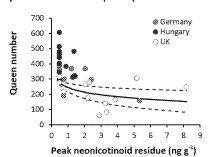
223	block/country'. † indicates where UK colony survival was too low for a formal analysis. Note
224	effect sizes differ between countries.
225	
226	Fig.3. Wild bee reproductive success in response to neonicotinoid nest residues. Separate
227	graphs are shown for the response of B. terrestris queen production and O. bicornis reproductive
228	cell production to neonicotinoid residues found in nests. The significance of these relationship is
229	based on a likelihood ratio test comparison of H0: ' $y \sim country$ ' and H1: ' $y \sim$
230	Neonicotinoid+country'. Neonicotinoid residues are based on summed concentrations of
231	clothianidin, thiamethoxam and imidacloprid. Expl. Var=Explained variance.
232	





ED_006569N_00037403-00011

a) Bombus terrestris queen production



b) Osmia bicornis reproductive cells

